

SAMPLE PREPARATION INSTRUCTIONS

Note: Cells should be spotted at a target density of 1 x 106 cells/mL on FTA™ paper. Cell counting is recommended as cells spotted at a density less than 0.8×10^6 cells/mL or greater than 1.7×10^6 cells/mL may not yield acceptable results.

- 1. Fill out a separate Cell Authentication Sample Submission Form for each sample submitted. Be sure the Barcode Number on the top of the Sample Submission Form matches the Barcode Number on the Sample Collection Card.
- 2. Prepare the samples one at a time at an optimal target cell density of 1×10^6 cells/mL.
 - a. For attached cells: Trypsinize and centrifuge at 125 x q. Discard the supernatant and resuspend the cell pellet in a small volume of PBS. Count the cells and dilute the sample to 1×10^6 cells/mL. If the cells are too dilute, re-centrifuge and resuspend them in a volume of PBS that will result in a spotting density of 1 x 10⁶ cells/mL.
 - b. For suspension cells: Harvest and count the cells. If cell density is greater than 1.7 x 106 cells/mL, dilute the sample in PBS to 1 x 106 cells/mL. If cell density is less than 0.8 x 106 cells/mL, centrifuge the cells and resuspend them in a volume of PBS that will result in a cell density of 1 x 106
- 3. Before handling the Sample Collection Card, thoroughly clean the work surface. With gloved hands, carefully open the Sample Collection Kit and remove the Sample Collection Card. Important: Wear gloves when handling the Sample Collection Cards to avoid cross-contamination with your own DNA.
- 4. Clearly label the Sample Collection Card with the cell line name/designation. If sending multiple cell lines, use a separate card for each cell line and make sure the information on the card matches the information on the Sample Submission Form.
- 5. Carefully mix and spot 40 µL of the cell suspension prepared in step 2 above at 1 x 10° cells/mL in the center of the circle on the inside of the Sample Collection Card.
- 6. Allow the Sample Collection Card to air dry in a laminar flow hood at room temperature (recommended drying time is at least 15 minutes).
- 7. When the Sample Collection Card is dry, place it and one desiccant pack (provided with the Kit) in the Multibarrier Pouch. Important: To avoid crosscontamination, use one Multibarrier Pouch per sample being submitted.
- Be sure to completely close the Multibarrier Pouch to preserve and protect the sample.
- 9. Repeat this process for each sample being submitted for testing, manipulating only one cell line at a time to avoid cross-contamination.
- 10. When the cell lines have been spotted and sealed in separate Multibarrier Pouches, place them and the completed corresponding Sample Submission Forms into the pre-addressed Return Envelope(s). If space allows, you may place multiple Multibarrier Pouches into one Return Envelope (be sure to include the Sample Submission Form(s)).
- 11. Affix the appropriate postage and place the sealed, pre-addressed Return Envelope(s) in the mail. If using an overnight service, please send the sample to the address listed on the pre-addressed Return Envelope. Important: For International Customers, please add the words "FTA Sample Papers" to the item description on your Air Waybill labels to avoid delays when mailing your samples.

Checklist

Before mailing your sample, did you?

- V Complete the Submission Form, be sure to complete the hazard statement
- ▼ Spot the cells in the Sample Card and place the card in the Ultra-barrier Pouch
- VIInclude the Submission Form and Multibarrier Pouch containing FTA Sample Card in the pre-addressed Return Envelope
- VI If mailing more than one sample, please be sure to match the FTA Card barcode number with the Submission Form barcode number
- ▼ Add words "FTA Sample Papers" to item description on Air Waybill labels
- V Mail the pre-addressed Return Envelope

KEEP THIS PORTION FOR YOUR RECORDS. Not For Medical Diagnostic Use.

Cell Authentication Service - Human STR Testing









SAMPLE SUBMISSION FORM

Place Barcode Label

Thank you for placing an online order for the Human STR Testing Service. Please read this form in its entirety and follow all steps accurately.

Peace of mind in 3 easy steps

	The STR Testing Results will be emailed by ATCC to the email address provided below. Customer Information (All fields are required. Please print information to ensure it is legible)		
Na	Name Koshio Funakoshi		
In	Institution/Company Funakoshi Co., Ltd. Address 9-7 Hongo 2-Chome		
Ad			
Ci	ity Bunkyo-ku State Tokyo Zip code 113-0033 Country Japan		
Er	mail ATCC@funakoshi.co.jp		
	(Please print clearly; authentication results will be sent to the email address above) $+81-3-5684-1645$ (Include Country Code)		
(Ir			
	Prepare your sample according to the Sample Preparation Instructions found on the back of this form. Enter the cell line information and sign the hazard statement below.		
H	Human Cell Line Information		
Pl	Please verify that the barcode number on your sample card matches the barcode number on this form for each sample.		
Н	Human Cell Line Name/Designation HEK-293 Catalog/Item # (If any) CRL-1573		
На	Hazard Statement		
	ATCC does not accept cultures infected with HIV or Biosafety Level 3 or 4 agents for STR testing. To the best of my knowledge the cell line being submitted is free of hazardous materials, agents, and carcinogens.		
Pr	rint Name Koshio Funakoshi		
Si	gnature Koshio Funakoshi Date MM/D	DD/YY	
3 M	ail your sample and this completed form in the pre-addressed Return Envelope included within the kit.		
IMPORTANT: For International Customers, please add the words " FTA Sample Papers " in the item description on your Air Wadelays when mailing your samples. If including more than one sample, please fill out a separate Cell Authentication Testing Service Submission Form for each sample submitted.			
KEEP TH	IIS PORTION FOR YOUR RECORDS		
Date sample submitted MM/DD/YY			
Name	Koshio Funakoshi _{E-mail} atcc@funakoshi.co.jp	Place Barcode Label	

STRTesting 0719-03

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